Impella CP or VA-ECMO in profound cardiogenic shock: left ventricular unloading and organ perfusion in a large animal model

Ole K. Møller-Helgestad1*, MD; Janus A. Hyldebrandt2, MD, PhD; Ann Banke1, MD; Charlotte S. Rud1, MD; Nanna L.J. Udesen1, MD; Louise Linde1, MD; Lisette O. Jensen1, MD, PhD, DMSc; Henrik Schmidt1, MD, DMSc; Hanne B. Ravn3, MD, PhD, DMSc; Jacob E. Møller1, MD, PhD, DMSc

1. Department of Cardiology, Cardiothoracic Surgery and Intensive Care, Odense University Hospital, Odense, Denmark; 2. Department of Anaesthesiology and Intensive Care, Akershus University Hospital, Lørenskog, Norway; 3. Heart Center, Copenhagen University Hospital, Copenhagen, Denmark

H.B. Ravn and J.E. Møller contributed equally as senior authors of this paper.

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Abstract

Aims: The aim of this study was to evaluate the Impella CP over veno-arterial extracorporeal membrane oxygenation (VA-ECMO) and their impact on left ventricular unloading and end-organ perfusion.

Methods and results: Cardiogenic shock (CS) was induced by injecting microspheres into the left coronary artery in fourteen adult female swine. Impella CP or VA-ECMO was initiated in the presence of CS and evaluated after 60 minutes. Left ventricular pressure-volume area (PVA, total mechanical work) was obtained from a conductance catheter. Results are presented as mean (95% confidence interval) and the rank-sum test was used to assess differences between devices. Compared to the CS state, PVA was unaffected by Impella CP and increased on VA-ECMO (from 2,548 [2,193; 2,904] mmHg x mL during CS to 5,775 [4,451; 7,099], between device p-value=0.02). Arterial lactate increased during CS and decreased on support with no difference between devices. Renal venous oxygen saturation decreased during CS and increased on support with no difference between devices. Cerebral venous oxygen saturation increased to 33% [25, 40] on Impella CP and to 69% [49, 89] on VA-ECMO, p=0.04.

Conclusions: In this porcine model of profound CS, Impella CP unloaded the left ventricle compared to VA-ECMO. Both devices improved end-organ perfusion, with a tendency towards higher venous oxygen saturations on VA-ECMO.

KEYWORDS

• acute heart failure
• cardiogenic shock
• ventricular assist device

*Corresponding author: Department of Cardiology, Odense University Hospital, Sdr. Boulevard 29, Entrance 20, 4th floor, 5000 Odense C, Denmark. E-mail: Ole.Moller-Helgestad@rsyd.dk
Introduction
The use of mechanical circulatory support (MCS) in cardiogenic shock (CS) has increased dramatically over the last 15 years. This was in an attempt to improve mortality in CS that continues to be approximately 50%1-3. Use of MCS in CS is currently based on expert opinion; guidelines only recommended MCS to be used in selected patients4,5. Two of the most frequently applied devices for patients admitted with CS are the Impella® CP (Abiomed, Danvers, MA, USA) and veno-arterial extracorporeal membrane oxygenation (VA-ECMO). However, there are no randomised controlled trials to provide evidence on which device or combination of devices, if any, provides optimal support. Current data based on retrospective cohort studies are difficult to interpret, with data suggesting the benefit of left ventricular unloading prior to revascularisation6-7. Further, VA-ECMO on top of an initial unloading device was recently associated with reduced mortality compared to when the two devices were applied simultaneously, emphasising the pertinent issue of improving end-organ perfusion versus ventricular unloading8. Also, patient groups comprise a heterogeneous population in terms of severity of CS and shock progression at the time of admission, impairing the ability to understand the potential benefits and adverse effects of the individual MCS device. This highlights the importance of testing the ability of these devices to provide both organ perfusion and unloading of the left ventricle (LV) under standardised conditions mimicking the clinical scenario of CS. Thus, the primary objective of this study was to compare the Impella CP and VA-ECMO at equal flow rates in a closed-chest large porcine model with profound CS and their effect on the LV as well as the effect on end-organ perfusion. Furthermore, this study also sought to describe the effect of maximising VA-ECMO flow.

Methods
A detailed description of anaesthesia and instrumentation can be found in Supplementary Appendix 1. Experiments were conducted with approval from and in accordance with guidelines from the Danish Animal Experiments Inspectorate (authorisation number: 2016-15-00951). Fourteen Danish Landrace female swine, approximately 18 weeks old and weighing 74.2±2.9 kg, were used.

LV PRESSURE-VOLUME MEASUREMENTS
A conductance catheter was inserted through a sheath in the right carotid artery and advanced retrogradely into the LV and connected to an MPVS Ultra® Pressure-Volume (PV) Loop System (Millar Inc., Houston, TX, USA). The MPVS Ultra was connected to a PowerLab 16/35 (ADInstruments, Dunedin, New Zealand) and PV measurements were continuously recorded in LabChart Pro (ADInstruments). Volumes were calibrated using an alpha correction9 and parallel wall conductance was determined using the hypertonic saline method. Data from the conductance catheter comprised: pressure-volume area (PVA, mmHg×mL), LV end-diastolic pressure (LVEDP, mmHg), LV end-diastolic volume (LVEDV, mL), LV stroke work (SW, mmHg×mL), LV output (mL/min and representing total cardiac output before and native LV output after MCS is initiated), LV end-systolic pressure-volume relationship (ESPVR, representing contractility), LV peak pressure and the LV isovolumic relaxation constant (Tau, representing ventricular relaxation). The gold standard for estimating ESPVR and PVA is by preload reduction, which was carried out at the study start but not when unsupported profound CS was achieved due to the severely compromised haemodynamics. Further, performing preload reductions during VA-ECMO support would have a major impact on device performance and thereby potentially alter device-derived afterload, making it difficult to extrapolate which component of the heart or device was responsible for changes in ESPVR and PVA. Therefore, baseline preload reductions were performed in each animal and the acquired Vo (the theoretical volume when no pressure is generated) was kept as a constant throughout the study to generate single-beat estimations of ESPVR and PVA (Figure 1)10,11. All other variables were derived automatically from the software.

EXPERIMENTAL PROTOCOL
The method for inducing CS has been described previously: in brief, 0.125 g polyvinyl alcohol microspheres (Contour™; Boston Scientific, Marlborough, MA, USA) mixed with 10 mL saline and 10 mL contrast was injected stepwise into the left main coronary artery through a JL 3.5 guide catheter (Launcher; Medtronic, Minneapolis, MN, USA) until the animal had developed profound CS, defined as cardiac output ≤2.0 L/min and/or a mixed venous oxygen saturation ≤35%, and MCS was initiated immediately thereafter12. The Impella CP was inserted through an introducer sheath in the left femoral artery, advanced retrogradely and placed with the inlet in the LV and outlet in the ascending aorta. The pump speed was set to P-8 and maintained for 60 minutes before withdrawing support. The venous cannula for the VA-ECMO was placed from the right femoral vein with the inlet just distal to the inferior vena cava ostium, and the arterial cannula was placed from the left femoral artery with the outlet in the left common iliac artery. Pump flow was set at 3.2 L/min for 60 minutes and then increased to maximum flow for 30 minutes before withdrawing support.

DATA ACQUISITION
Arterial as well as cerebral and renal venous blood gases for oxygen saturation and lactate measurements were sampled at baseline, when CS had been induced, and after 60 minutes of circulatory

Abbreviations
ESPVR  end-systolic pressure-volume relationship
LV  left ventricle
LVEDP  left ventricular end-diastolic pressure
LVEDV  left ventricular end-diastolic volume
MAP  mean arterial pressure
MCS  mechanical circulatory support
PV  pressure-volume
PVA  pressure-volume area
SW  left ventricular stroke work
VA-ECMO  veno-arterial extracorporeal membrane oxygenation
support as well as 30 minutes after maximising VA-ECMO flow. PV relationships were determined at baseline using preload reduction and during shock and MCS manually selecting loops keeping \( V_0 \) (theoretical volume when no pressure is generated) attained from baseline as a constant. A) Animal receiving Impella CP. B) Animal receiving VA-ECMO. PE: potential energy, internal energy; SW: stroke work, external energy; PVA is the total mechanical work done by the ventricle per heartbeat and is the sum of PE and SW.

**Figure 1.** Examples of how the end-systolic (ESPVR, red line) and end-diastolic (EDPVR, light blue line) PV relationships were determined at baseline using preload reduction and during shock and MCS manually selecting loops keeping \( V_0 \) (theoretical volume when no pressure is generated) attained from baseline as a constant. A) Animal receiving Impella CP. B) Animal receiving VA-ECMO. PE: potential energy, internal energy; SW: stroke work, external energy; PVA is the total mechanical work done by the ventricle per heartbeat and is the sum of PE and SW.

Statistical analyses were performed with Stata/IC 14 (StataCorp, College Station, TX, USA). A p-level \( \leq 0.05 \) was considered statistically significant.

**Results**

**BASELINE CHARACTERISTICS**

CS was successfully induced in all animals. One animal developed intractable ventricular fibrillation two minutes after initiating VA-ECMO and was excluded from the study. Another animal had invalid volume signals from the PV catheter and was also excluded. Two minutes after maximising VA-ECMO flow, one further animal developed irreversible ventricular fibrillation; therefore, data on maximum VA-ECMO flow are reported in four animals.

Animals received 13 [10, 17] mL and 15 [9, 20] mL microsphere solution, \( p=0.75 \), in the Impella CP and VA-ECMO group, respectively, and there were no differences between groups in the measured variables at baseline or when CS was induced (Table 1, Table 2). One animal in the VA-ECMO group and all animals in the Impella CP group required norepinephrine 0.01-0.2 µg/kg/min to maintain a mean arterial pressure (MAP) >40 mmHg after CS.
was established. The one animal in the VA-ECMO group was able to be successfully weaned off norepinephrine shortly after VA-ECMO had been initiated, whereas all animals on Impella CP required norepinephrine throughout the mechanical support. One animal in each group was successfully resuscitated from asystole occurring immediately before initiating circulatory support. Impella CP flow was 3.1 [3.0, 3.2] L/min, borderline significantly lower than the VA-ECMO group: 3.2 [3.2, 3.3] L/min, p=0.05. A flow of 4.6±0.2 L/min was achieved when maximising VA-ECMO flow. Except for one animal in each group, complete circulatory collapse with pulseless electrical activity developed within a few minutes after withdrawal of MCS.

**Table 1. Haemodynamics, lactate levels and venous oxygen saturations at different time points.**

<table>
<thead>
<tr>
<th></th>
<th>Baseline mean (95% CI)</th>
<th>p-value</th>
<th>Cardiogenic shock mean (95% CI)</th>
<th>p-value</th>
<th>60 minutes support mean (95% CI)</th>
<th>p-value</th>
<th>90 minutes VA-ECMO support mean (95% CI)</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>MAP, systemic (mmHg)</td>
<td>Impella CP, n=6</td>
<td>64 [56, 71]</td>
<td>0.87</td>
<td>31 [26, 35]</td>
<td>0.47</td>
<td>54 [45, 63]</td>
<td>0.13</td>
<td>n=5 83 [72, 93]</td>
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<td>VA-ECMO, n=6</td>
<td>65 [56, 74]</td>
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<tr>
<td>Arterial lactate (mmol/L)</td>
<td>Impella CP, n=6</td>
<td>2.1 [1.7, 2.6]</td>
<td>0.06</td>
<td>3.8 [2.9, 4.7]</td>
<td>0.26</td>
<td>1.7 [1.1, 2.3]</td>
<td>0.75</td>
<td>n=4 2.3 [0.8, 5.3]</td>
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<td>VA-ECMO, n=6</td>
<td>1.4 [0.9, 1.8]</td>
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<tr>
<td>Venous lactate, cerebral (mmol/L)</td>
<td>Impella CP, n=6</td>
<td>3.8 [2.0, 5.6]</td>
<td>0.30</td>
<td>6.3 [4.1, 8.4]</td>
<td>0.09</td>
<td>3.9 [2.7, 5.1]</td>
<td>0.85</td>
<td>n=4 4.0 [0.1, 7.9]</td>
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<td>VA-ECMO, n=6</td>
<td>2.6 [1.6, 3.6]</td>
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<tr>
<td>Venous lactate, renal (mmol/L)</td>
<td>Impella CP, n=5</td>
<td>2.2 [1.7, 2.7]</td>
<td>0.08</td>
<td>2.7 [1.1, 4.3]</td>
<td>0.86</td>
<td>2.3 [0.4, 4.1]</td>
<td>0.78</td>
<td>n=4 1.5 [0.1, 3.0]</td>
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<tr>
<td>VA-ECMO, n=6</td>
<td>1.5 [1.0, 2.0]</td>
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<td>Venous O2 sat cerebral (%)</td>
<td>Impella CP, n=6</td>
<td>51 [36, 67]</td>
<td>0.75</td>
<td>20 [17, 23]</td>
<td>0.17</td>
<td>33 [25, 40]</td>
<td>0.04</td>
<td>n=4 65 [28, 101]</td>
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<tr>
<td>VA-ECMO, n=6</td>
<td>58 [52, 63]</td>
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<tr>
<td>Venous O2 sat renal (%)</td>
<td>Impella CP, n=5</td>
<td>86 [78, 93]</td>
<td>0.57</td>
<td>50 [25, 75]</td>
<td>0.14</td>
<td>59 [34, 84]</td>
<td>0.10</td>
<td>n=4 80 [54, 105]</td>
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<td>VA-ECMO, n=6</td>
<td>87 [84, 90]</td>
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<td>Heart rate (beats per minute)</td>
<td>Impella CP, n=6</td>
<td>69 [61, 77]</td>
<td>0.22</td>
<td>68 [62, 74]</td>
<td>0.47</td>
<td>87 [73, 100]</td>
<td>0.03</td>
<td>n=5 78 [54, 102]</td>
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<tr>
<td>VA-ECMO, n=6</td>
<td>76 [67, 84]</td>
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</table>

Data are presented as mean [95% confidence intervals] and n indicates sample size.

**Table 2. LV PV variables at different time points.**

<table>
<thead>
<tr>
<th></th>
<th>Baseline mean (95% CI)</th>
<th>p-value</th>
<th>Cardiogenic shock mean (95% CI)</th>
<th>p-value</th>
<th>Immediate response, mean (95% CI)</th>
<th>p-value</th>
<th>60 minutes support mean (95% CI)</th>
<th>p-value</th>
<th>90 minutes VA-ECMO support mean (95% CI)</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>LVEDV (mL)</td>
<td>Impella CP, n=6</td>
<td>153 [140, 166]</td>
<td>0.30</td>
<td>182 [152, 213]</td>
<td>0.08</td>
<td>119 [65, 173]</td>
<td>0.007</td>
<td>114 [78, 151]</td>
<td>0.03</td>
<td>n=5 170 [122, 219]</td>
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<tr>
<td>VA-ECMO, n=6</td>
<td>143 [133, 152]</td>
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<td>147 [112, 181]</td>
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<tr>
<td>LVEDP (mmHg)</td>
<td>Impella CP, n=6</td>
<td>14 [12, 17]</td>
<td>0.57</td>
<td>20 [17, 22]</td>
<td>0.30</td>
<td>14 [9, 19]</td>
<td>0.004</td>
<td>13 [10, 15]</td>
<td>0.007</td>
<td>n=5 19 [14, 24]</td>
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<tr>
<td>VA-ECMO, n=6</td>
<td>16 [12, 19]</td>
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<td>20 [14, 25]</td>
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<tr>
<td>PVA (mmHg×mL)</td>
<td>Impella CP, n=6</td>
<td>6,555 [5,637, 7,073]</td>
<td>0.75</td>
<td>2,792 [2,296, 3,287]</td>
<td>0.26</td>
<td>2,456 [1,557, 4,356]</td>
<td>0.004</td>
<td>3,129 [1,616, 4,663]</td>
<td>0.02</td>
<td>n=5 8,030 [7,146, 8,914]</td>
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<tr>
<td>VA-ECMO, n=6</td>
<td>6,780 [5,445, 8,114]</td>
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<tr>
<td>LV peak pressure (mmHg)</td>
<td>Impella CP, n=6</td>
<td>85 [77, 93]</td>
<td>0.63</td>
<td>37 [29, 44]</td>
<td>0.87</td>
<td>46 [31, 60]</td>
<td>0.004</td>
<td>66 [49, 84]</td>
<td>0.15</td>
<td>n=5 102 [80, 125]</td>
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<td>VA-ECMO, n=6</td>
<td>84 [74, 93]</td>
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<td></td>
<td>86 [61, 111]</td>
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<tr>
<td>SW (mmHg×mL)</td>
<td>Impella CP, n=6</td>
<td>3,826 [3,432, 4,220]</td>
<td>0.52</td>
<td>478 [354, 741]</td>
<td>0.52</td>
<td>813 [341, 1,409]</td>
<td>0.42</td>
<td>1,337 [341, 2,332]</td>
<td>1.00</td>
<td>n=5 1,365 [35, 2,659]</td>
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<tr>
<td>VA-ECMO, n=6</td>
<td>3,806 [2,683, 4,930]</td>
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<td>1,265 [349, 2,182]</td>
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<tr>
<td>LV output (mL/min)</td>
<td>Impella CP, n=6</td>
<td>3,198 [3,17, 2,689]</td>
<td>0.75</td>
<td>2,313 [1,787, 2,839]</td>
<td>0.63</td>
<td>3,146 [2,573, 3719]</td>
<td>1.00</td>
<td>3,685 [2,138, 5,331]</td>
<td>0.26</td>
<td>n=5 2,754 [2,610, 3,908]</td>
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<tr>
<td>VA-ECMO, n=6</td>
<td>5,182 [4,548, 5,815]</td>
<td></td>
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<td>2,812 [1,873, 3,751]</td>
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<tr>
<td>ESPVR (mmHg/mL)</td>
<td>Impella CP, n=6</td>
<td>1.08 [0.84, 1.31]</td>
<td>0.20</td>
<td>0.31 [0.25, 0.37]</td>
<td>0.34</td>
<td>0.44 [0.27, 0.61]</td>
<td>0.04</td>
<td>1.09 [0.61, 1.58]</td>
<td>0.26</td>
<td>n=5 0.88 [0.50, 1.25]</td>
</tr>
<tr>
<td>VA-ECMO, n=6</td>
<td>1.31 [1.07, 1.55]</td>
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<td></td>
<td></td>
<td>0.91 [0.54, 1.27]</td>
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<tr>
<td>Tau (ms)</td>
<td>Impella CP, n=6</td>
<td>42 [35, 50]</td>
<td>0.69</td>
<td>133 [76, 190]</td>
<td>0.15</td>
<td>153 [92, 214]</td>
<td>0.20</td>
<td>52 [35, 69]</td>
<td>0.01</td>
<td>n=5 76 [59, 94]</td>
</tr>
<tr>
<td>VA-ECMO, n=6</td>
<td>40 [35, 45]</td>
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<td></td>
<td>89 [62, 116]</td>
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n indicates sample size. LV output: total cardiac output before MCS, native left ventricular output while on support; Tau: left ventricular isovolumic relaxation constant.
END-ORGAN PERFUSION
Both devices improved MAP although the need for norepinephrine was greater for Impella animals. Markers of end-organ perfusion improved in both groups, with a trend towards greater improvement in regional venous oxygen saturation on VA-ECMO support (Table 1). Maximising VA-ECMO seemed to increase MAP but had no effect on markers of end-organ perfusion.

LV UNLOADING
PV data are summarised in Table 2 and the repeated measurements of LVEDV and LVEDP throughout the study period can be found in Figure 2 and Figure 3, respectively. After 60 minutes of MCS, heart rate was significantly higher in the Impella CP group (Table 2). MCS resulted in an immediate decrease in LVEDV and LVEDP in the Impella CP group compared to a tendency to increased LVEDV and an increase in LVEDP in the VA-ECMO group (Figure 4), resulting in a significantly lower PVA in the Impella CP group. These findings were maintained during the initial 60 minutes of circulatory support, during which the diastolic relaxation constant, Tau, only improved in the Impella group. Active SW and native LV volume output showed a tendency towards being increased in both groups after initiating MCS with no statistical difference between groups.

Discussion
Circulatory support with Impella CP and norepinephrine more efficiently unloaded the LV compared to peripheral VA-ECMO in a large closed-chest porcine model of profound CS. Both strategies

**Figure 2.** LVEDV over time. Dots represent mean values and error bars standard error of the mean.

**Figure 3.** LVEDP over time. Dots represent mean values and error bars standard error of the mean.
improved organ perfusion by lactate clearance and increased cerebral/renal venous oxygen saturation, with a tendency towards higher venous oxygen saturation on VA-ECMO support.

**LV UNLOADING**

Theoretically, Impella CP decreases and VA-ECMO increases LV loading and the loading by VA-ECMO is expected to be greater with higher flow, which is why the study was designed to compare the devices at equal flow rates. PVA was chosen as the primary outcome, representing total mechanical (pressure and volume) work per heartbeat and linearly correlated with myocardial oxygen consumption independent of heart rate\(^{13}\). PVA is the sum of all mechanical energy performed by the ventricle per heartbeat and comprises external active forces, stroke work (SW) and internal passive forces, potential energy (PE) as shown in **Figure 1A** within the Impella CP PV example. Comparing systemic cardiac output during MCS is a challenge, as standard monitoring using a Swan-Ganz catheter in the pulmonary artery is not applicable when VA-ECMO almost completely bypasses the pulmonary circulation. For this reason, organ perfusion was evaluated using lactate levels given their relationship with oxygenation\(^{14}\) and regional venous oxygen saturations given their relationship with blood flow applying the Fick principle.

After an immediate increase in LVEDP and LVEDV in the VA-ECMO group, the LV seemed to adapt as LVEDP and LVEDV gradually decreased over the 60 min (**Figure 1**). Interestingly, when maximising VA-ECMO flow, LVEDV did not adapt to the same degree as during less flow (**Figure 1**), and PVA was markedly increased without any real changes in markers of tissue perfusion, meaning that the main effect of maximising VA-ECMO flow was an increased PVA. In accordance with our findings, a porcine study of CS reported that increasing VA-ECMO flow showed tendencies towards increased LVEDV\(^{15}\), but the study did not investigate changes in LVEDV over time as they increased VA-ECMO flow every five minutes. It is difficult to say whether the observed LVEDV adaption during low VA-ECMO flow was due to reductions in systemic vascular resistance, reduction in preload or better contractility in non-ischaemic myocardium. The lack of adaption during full flow speaks against a reduction in preload. The systemic vascular resistance component of afterload is difficult to calculate as total cardiac output during VA-ECMO is difficult to determine accurately and it is likely that the device-related component of afterload was constant since device flow was constant throughout the study. However, trends in LV contractility (ESPVR, **Table 2**) follow the LVEDV changes in the VA-ECMO animals, indicating that LVEDV adaption during low flow could be due to revived non-ischaemic myocardium.

**COMPARING STUDIES OF IMPELLA AND VA-ECMO**

To our knowledge, only two studies have compared MCS with the Impella and VA-ECMO head to head\(^{16,17}\) and our study is the first to compare the devices during CS due to LV failure. Kawashima et al compared the Impella LD\(^{8}\) (Abiomed), requiring surgical insertion with a maximal flow of 5 L/min, to VA-ECMO at comparable device flow rates in dogs weighing 20 to 24 kg\(^{16}\). They created acute LV failure after sternotomy in dogs with stepwise total LAD occlusion starting distally and moving proximally, testing the devices at each stage. Their study showed that the Impella LD unloads LV more than VA-ECMO, while maintaining systemic blood pressure. Ostadal et al compared the Impella 2.5\(^{8}\) (Abiomed) to VA-ECMO at different pump speeds targeting a MAP >70 mmHg in adult female swine weighing 50 to 60 kg\(^{17}\). They created different degrees of LV dysfunction by ventricular pacing from 200 beats/min to ventricular fibrillation. Their study showed that VA-ECMO was superior...
to the Impella 2.5 during the most severe heart failure condition in terms of maintaining adequate MAP, and that norepinephrine had to be initiated in the Impella 2.5 group to keep MAP ≥40 mmHg during ventricular fibrillation. In agreement with these studies we found that the Impella CP unloads the LV compared to VA-ECMO and that VA-ECMO generates a higher MAP. As opposed to the study by Kawashima et al16 where cardiac output only decreased from 2.46 to 2.08 L/min in animals weighing on average 75 kg. This could explain why Kawashima et al16 found no difference between the Impella LD and VA-ECMO in terms of systemic blood pressure. Another explanation could be that the Impella LD is powerful enough to maintain systemic blood pressure in severe heart failure, especially in smaller animals.

END-ORGAN PERFUSION

Successful treatment of CS involves breaking the vicious cycle of myocardial ischaemia, myocardial dysfunction, hypotension and low cardiac output14, and restoring organ perfusion, evident by improved survival in patients clearing arterial lactate21. In our study, LV contractility, defined as ESPVR, improved during MCS in both groups whereas the diastolic parameter Tau only improved in the Impella CP group. Systemic blood pressure and organ perfusion improved in both groups but at the expense of LV loading by higher LVEDV, LVEDP and PVA and thereby oxygen consumption in the VA-ECMO group. This indicates that the Impella CP with norepinephrine and VA-ECMO can improve organ perfusion and LV functions even during severe ischaemia, and that a combination of the Impella CP and norepinephrine does so without increasing myocardial oxygen consumption, which might protect the myocardium against further ischaemic injury30. The fact that all animals treated with the Impella CP required norepinephrine to maintain a MAP >40 mmHg is probably due to the Impella CP being an axial pump that, by design, increases flow but has limited impact on pressure, whereas VA-ECMO is a centrifugal pump that, by design, increases both flow and pressure.

Limitations

A limitation to our study is that we infused microspheres without a proximal occluding balloon, introducing the possibility that some microspheres affected end organs although a minimal effect can be assumed as oxygenation and lactate levels improved during support. Also, the model does not allow the assessment of device impact after reperfusion, and is unsuitable to assess infarct size, as the microcirculation is obstructed. Another limitation is the short ascending aorta of pigs, resulting in the Impella CP outflow being positioned in the aortic arch-descending aorta in at least one animal, possibly limiting device-related perfusion of the cerebrum in affected animals when using a femoral access approach. The seemingly higher arterial and cerebral venous lactate level on VA-ECMO support was driven by a single outlier with coherent low oxygen saturations, possibly due to hypocapnia as arterial pCO2 ranged between 12 and 18 mmHg during the first 60 minutes of MCS in that animal. This is a study on the immediate short-term effect of the Impella CP and VA-ECMO and it is unknown if the loading effects would become exaggerated with long-term MCS. Further, norepinephrine was used in this study, and it is unknown if other vasoactive agents have the same or a different impact on LV loading and end-organ perfusion in combination with the Impella CP. Also, Impella and VA-ECMO are sometimes used in combination when treating CS patients21, but so far it is unknown how the devices in combination affect the LV and end-organ perfusion, indicating the need for future studies.

Conclusions

In this closed-chest porcine model with profound CS due to microsphere injections into the left main coronary artery, a combination of the Impella CP and norepinephrine provided unloading of the LV compared to VA-ECMO. Both devices improved markers of end-organ perfusion, with a trend towards higher local venous oxygen saturations in the VA-ECMO group.

Impact on daily practice

Increasing end-organ perfusion without increasing myocardial oxygen demand is a key element in the treatment of CS but is often a double-edged sword. We demonstrated that both the Impella CP and VA-ECMO are capable of improving end-organ perfusion and the Impella CP does so without increasing myocardial oxygen demand. VA-ECMO might be more powerful in improving organ perfusion, but at the expense of a substantial increase in myocardial oxygen demand. These considerations should be taken into account when MCS is required.

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Conflict of interest statement

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Supplementary data
Supplementary Appendix 1. Anaesthesia.

The supplementary data are published online at: http://www.pcronline.com/eurointervention/148th_issue/271
Supplementary data

Supplementary Appendix 1. Anaesthesia

The fourteen swine were acquired from a licensed breeder and acclimatised for at least 3 days. On the day of the experiment the animal was premedicated with a combination of 0.2 mg/kg midazolam, 0.04 mg/kg medetomidine and 0.05 mg/kg atropine and general anaesthesia was induced by 5 mg/kg propofol and the animal was intubated. Anaesthesia was maintained with 1.8-3.1% isofluorane and a continuous infusion of 250-500 µg/hour fentanyl. During VA-ECMO support anaesthesia was supplemented with infusion of 10 mg/hour midazolam as pulmonary perfusion was considered too low to maintain anaesthesia with isofluorane alone. To avoid clotting of the catheters 20,000 IU of unfractionated heparin were injected intravenously every 2 hours. Norepinephrine was infused if needed during mechanical circulatory support to maintain MAP ≥ 40 mmHg. 300 mg of amiodarone was infused over 30 minutes, followed by continuous infusion of 60 mg/hour to avoid arrhythmias. After termination of the studies, animals were euthanised by a lethal dose of pentobarbital.

Instrumentation:

All animals were mechanically ventilated (MCM 801 Ventilator; Dameca A/S, Denmark) with a mixture of room air and oxygen to maintain normal oxygen saturation. Tidal volume and respiratory rate were adjusted to achieve a paCO₂ of 35-45 mmHg attained from serial blood gas analyses (epoc blood analysis; Epocal inc., Ottawa, ON, Canada). Surface electrocardiography leads, peripheral venous catheters, and a urinary bladder catheter were placed in all animals. Heating pads were used to maintain normothermia of 38-38.5°C. Vascular access sheaths were deployed by surgical cutdown in the neck and ultrasound-guided Seldinger technique in the groin. The left carotid artery was catheterised with a 6 Fr sheath and a JL3.5 guide catheter (Launcher; Medtronic Inc., Minneapolis, MN, USA) was placed in the left coronary. The left internal jugular vein was catheterised with a 4 Fr, 5 cm double lumen central line (Arrow International Inc, Reading, PA, USA) placed with the tip in the sigmoid sinus inside the skull for measurement of cerebral venous oxygen saturation and lactate levels. The right carotid artery was catheterised with an 8 Fr sheath and a conductance catheter (Venti-Cath 512 PV Loop Catheter; Millar Inc., Houston, TX, USA) was placed in the LV for
continuous recordings of pressure-volume (PV) relationships. The right external jugular vein was catheterised with a two-lumen central venous access device (MAC™; Arrow International Inc., Reading, PA, USA) for fluid administration. The left femoral vein was catheterised with a 10 Fr sheath and a Swan-Ganz catheter was advanced through the sheath to one of the renal veins with fluoroscopic guidance for measurement of renal venous oxygen saturation and lactate levels. The right femoral artery was catheterised with a 6 Fr sheath for measurement of systemic blood pressure and obtainment of arterial blood gases. The left femoral artery was catheterised with a 6 Fr sheath for easy access of the Impella introducer sheath or arterial cannula for the VA-ECMO. The right femoral vein was catheterised with a 14 Fr sheath and a balloon occlusion catheter was placed in the inferior vena cava at the level of the diaphragm (NUCLEUSTM; NuMED Canada Inc., Cornwall, ON, Canada) for preload reductions at baseline and later easy access of the venous cannula for the VA-ECMO.